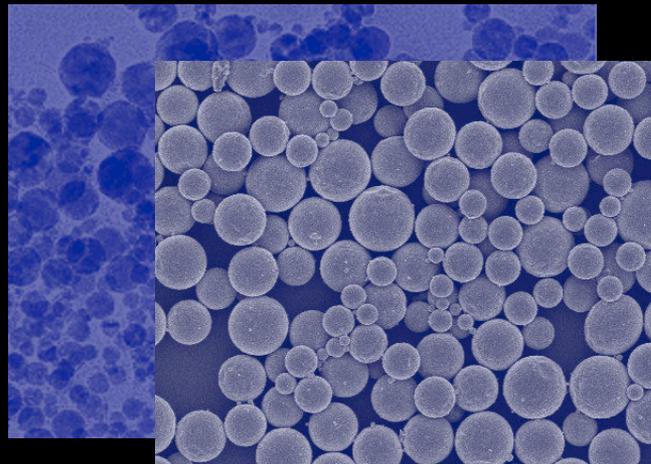


# Toxicity Evaluation of HfO<sub>2</sub> Nanoparticles

Scott Boitano<sup>1</sup>, Reyes Sierra<sup>2</sup>, Jim Field<sup>2</sup>, Buddy Ratner<sup>3</sup>, Farhang Shadman<sup>2</sup>

## Measuring cytotoxicity of nanoparticles in human cells



<sup>1</sup> Arizona Respiratory Center & Dept. of Physiology  
and Dept Chemical & Environmental Engineering<sup>2</sup>, University of Arizona

<sup>3</sup> University of Washington Engineered Biomaterials (UWEB)



U W E B

# HfO<sub>2</sub> Model Nanoparticles

- **Reference “Micron-sized” HfO<sub>2</sub> particles**
  - Reported particle size: < 44 μm;
  - Measured size (Laser Diffraction and Electron Microscopy; Sierra Group) showed wide range (200 nm-20 μm) with distribution peaks at 500 nm and 6 μm
- **Batch 1 nano-sized HfO<sub>2</sub> particles: “Batch-1 360”**
  - Reported particle size: ~20 nm
  - Measured size showed range of 60 nm - 1 μm with a peak at 360 nm
- **Batch 2 nano-sized HfO<sub>2</sub> particles: “Batch-2 224”**
  - Reported particle size: ~1-2 nm
  - Measured size showed range of 90 nm - 2 μm with a major peak at 224 nm
- **Batch 3 nano-sized HfO<sub>2</sub> particles: “Batch-3 169”**
  - Reported average particle size: ~100 nm
  - Measured size showed range of 70 - 400 nm with a peak at 169 nm

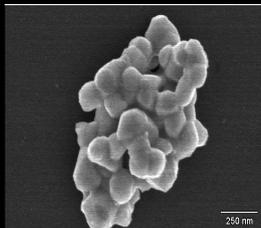
# Mammalian Cell Models: Epithelium

- We consider two major routes of exposure:
  - Lung (via inhalation or systemic exposure)
  - Skin (via direct contact with nanoparticles)
- Both are lined by epithelial cells that form a barrier to keep outside exposures from underlying cells and tissue
- We use two epithelial cell models for preliminary studies:
  - Human bronchial epithelial cell line (16HBE14o- cells)
  - Human skin epithelial cell line (HaCat)

# Measurements of Cytotoxicity with Fluorescence: Live/Dead Assay

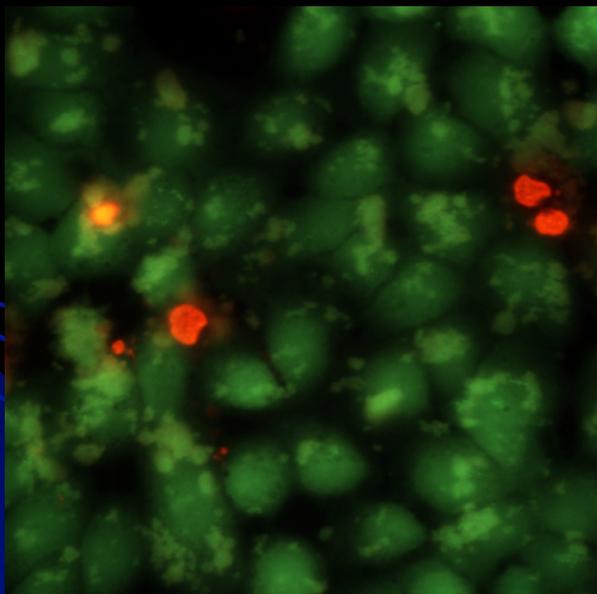
- Cells are incubated with  $\text{HfO}_2$  for 2 hr to induce toxicity
- Cells are then exposed to two low-fluorescent compounds: **Ethidium homodimer (EtHD-1)** and **Calcein-AM**
- **EtHD-1** can only enter damaged cells, where it interacts with DNA and increases its fluorescence 40x (ex/em ~495 nm/~635 nm)
- **Calcein-AM** can cross the plasma membrane of live cells. Once inside, the acetyl-methoxy ester groups (AM) are cleaved by non-specific cell esterases, resulting in charged and highly fluorescent molecules, **Calcein** (ex/em ~495 nm/~515 nm)
- Net result: In tact, **Live**, cells fluoresce **Green** and **Dead** cells fluoresce **Red**

# Live/Dead with Micron-sized HfO<sub>2</sub>

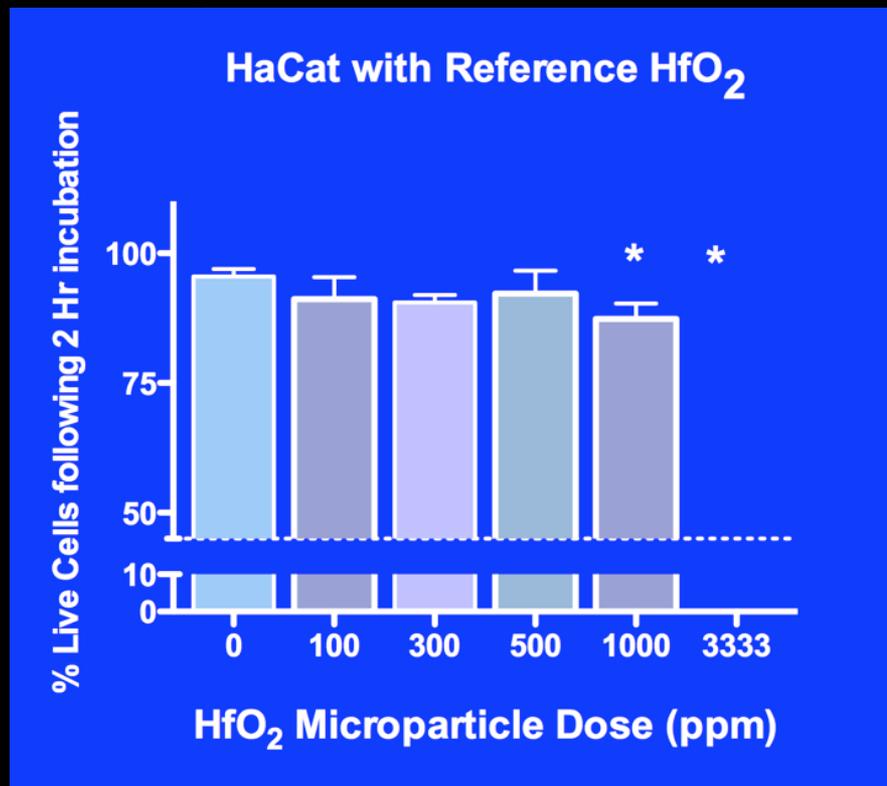


Major size peaks at 500 nm and 6 μm

SEM

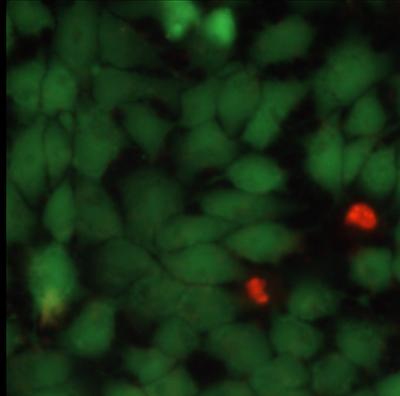
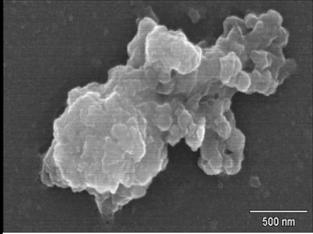


Live-Dead Fluorescence  
(1000 ppm)

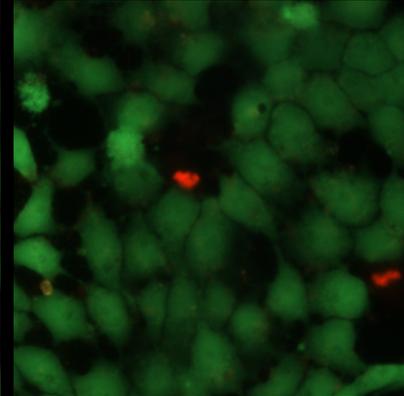


- Slight cytotoxicity at 1000 ppm
- Strong cytotoxicity at 3000 ppm
- 16HBE14o- (lung) epithelial cytotoxicity was similar

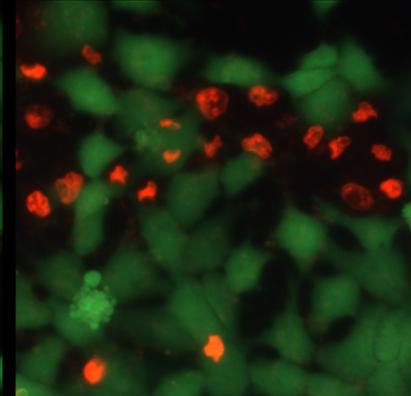
# Live/Dead with Batch-1 360 HfO<sub>2</sub>



50 ppm

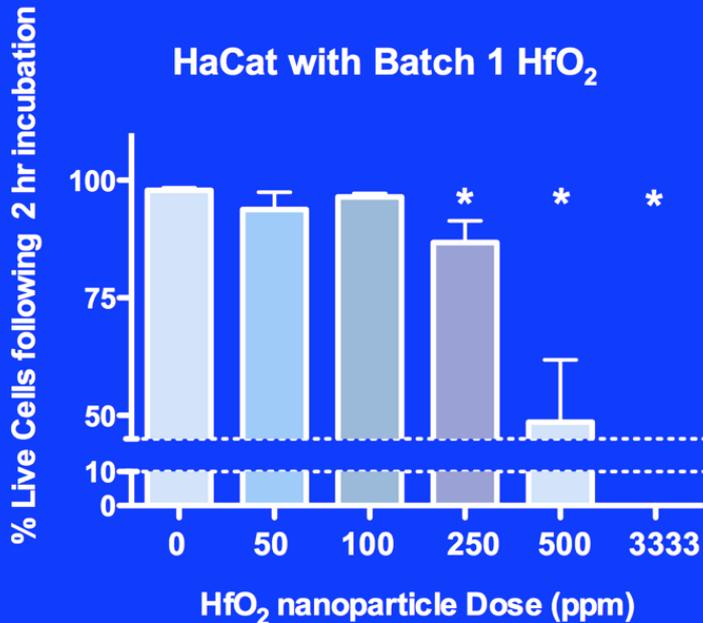


250 ppm



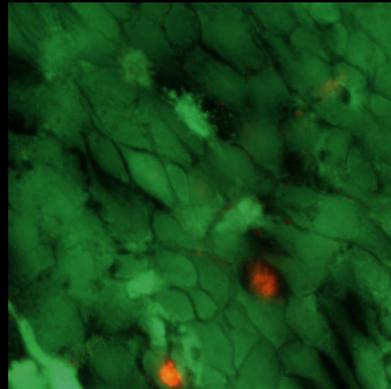
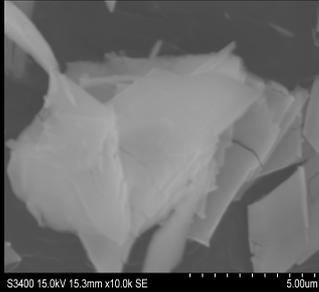
500 ppm

HaCat with Batch 1 HfO<sub>2</sub>

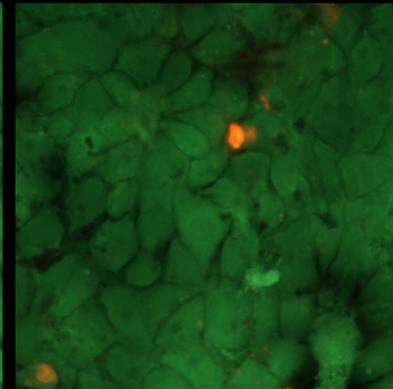


- Slight cytotoxicity at 250 ppm
- Strong cytotoxicity at 500 ppm
- Smaller HfO<sub>2</sub> particles are more toxic to HaCat (skin) epithelial cells
- 16HBE14o- (lung) epithelial cells were similar

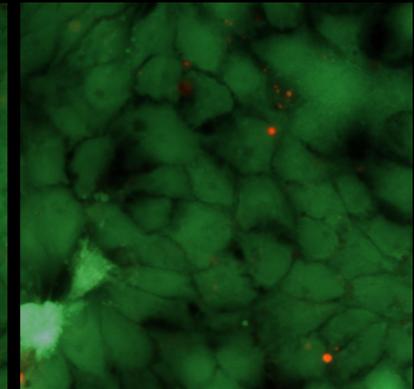
# Live/Dead with Batch-2 224 HfO<sub>2</sub>



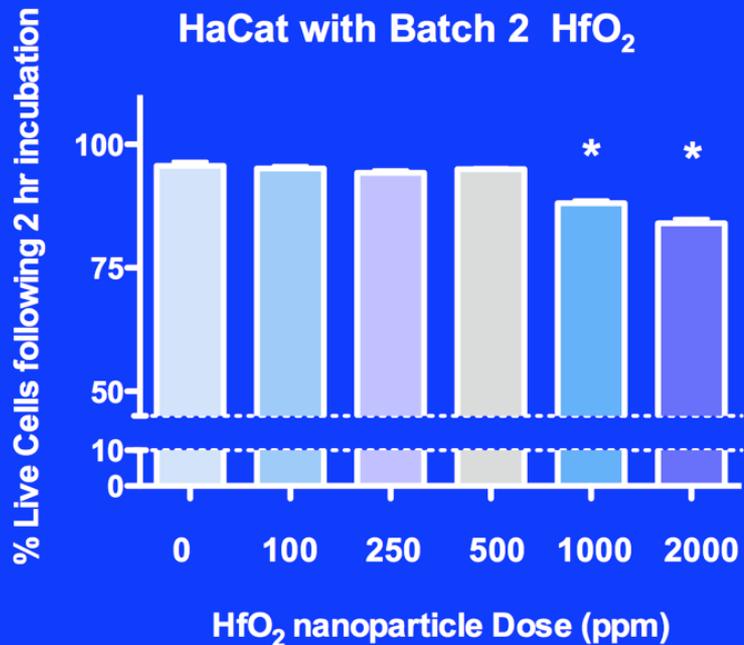
50 ppm



250 ppm

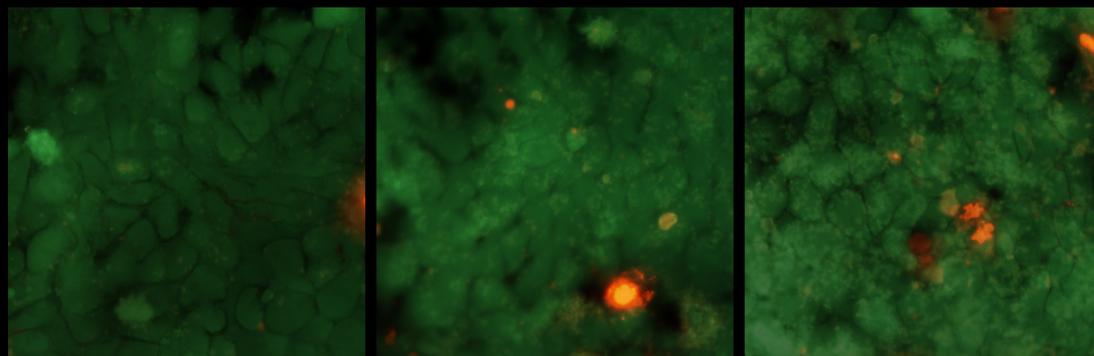
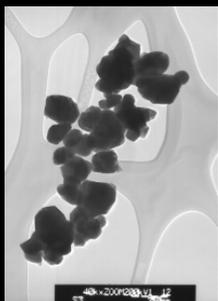


500 ppm



- Slight cytotoxicity at 1000 ppm and at 2000 ppm
- Although Batch-2 224 contain smaller particles than Batch-1 336, less cytotoxicity is observed in HaCat (skin) epithelial cells
- 16HBE14o- (lung) epithelial cells were similar

# Live/Dead with Batch-3 169 HfO<sub>2</sub>

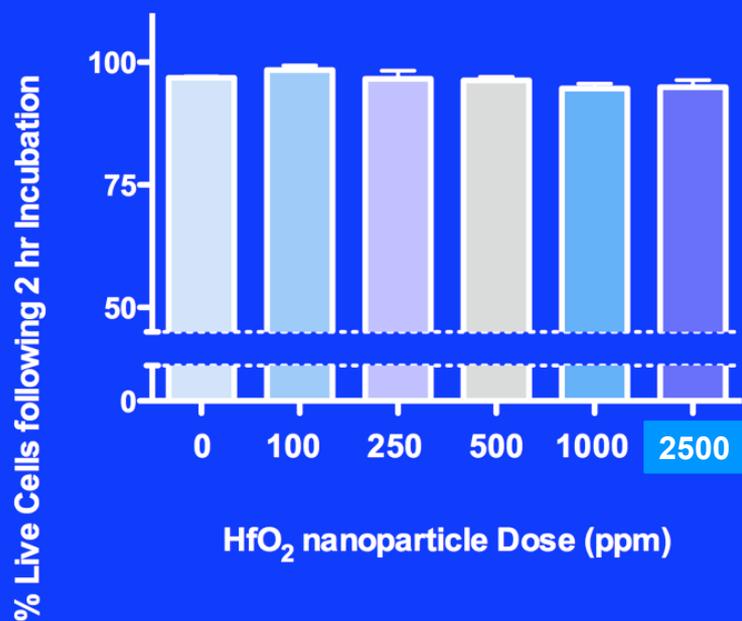


250 ppm

1000 ppm

2500 ppm

HaCat with Batch 3 HfO<sub>2</sub>



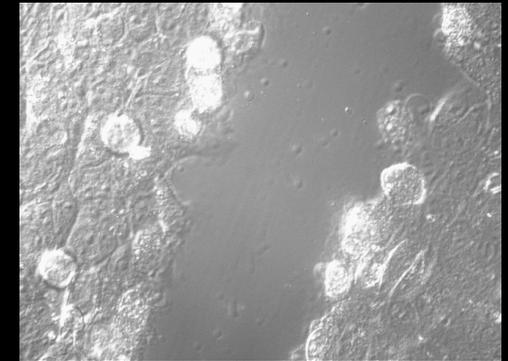
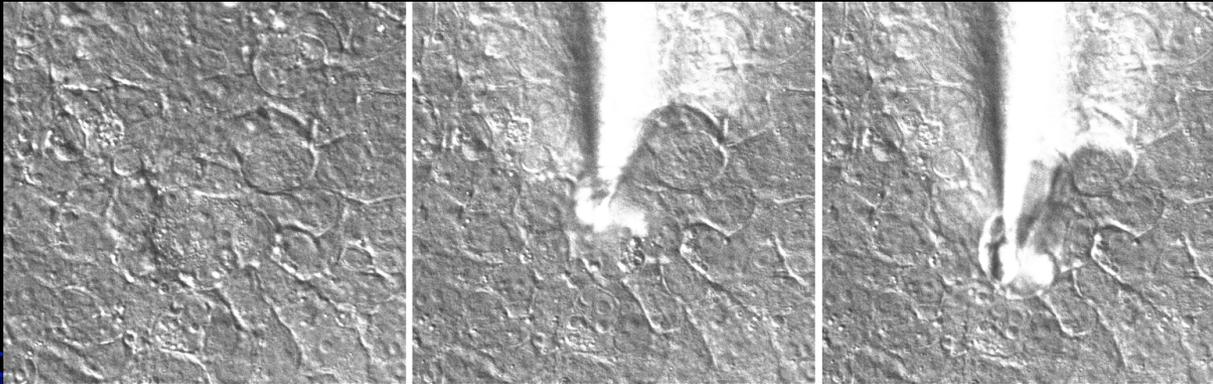
- No cytotoxicity up to 2500 ppm
- HfO<sub>2</sub> particles with the smallest peak size displayed the least cytotoxicity in HaCat (skin) epithelial cells
- 16HBE14o- (lung) epithelial cells not tested

# Live-Dead Cytotoxicity Summary

- Live-Dead assay is a straight forward measure for cytotoxicity that works well with mammalian cells
- Similar to Microtox and MTT Assays from Sierra Group, Batch-1 360 HfO<sub>2</sub> displayed highest cytotoxicity
- Smaller HfO<sub>2</sub> sizes did not correlate directly with increased toxicity
- SIMS measurements (Ratner Group) showed different contaminants between Batch-1 360 and Batch-2 224 -- might the contaminants be important in cytotoxicity?
- Live-Dead assay is, however, limited:
  - Necessary to choose a single time point for analysis
  - Requires plasma membrane disruption and thus may not reflect HfO<sub>2</sub>-induced cell compromise

# Scrape Wound Assay to Evaluate Cytotoxicity

- Drag a glass pipet across an epithelial monolayer (16HBE14o- cells used for these studies)



- Follow re-establishment of the monolayer using video microscopy
- We used this technique to evaluate if  $\text{HfO}_2$  particles altered wound repair

# Batch-1 360 HfO<sub>2</sub> Inhibits Wound Repair

Note: MPG videos of each repair are available via download

0 ppm Time: 0 hr



0.5 hr



1.0 hr



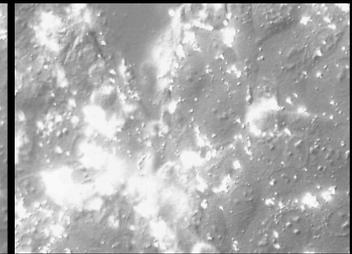
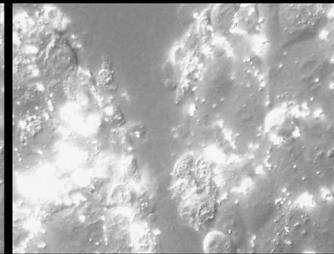
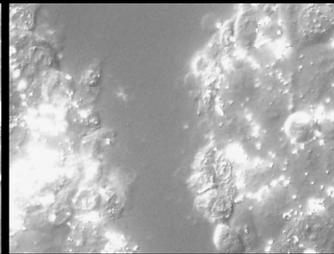
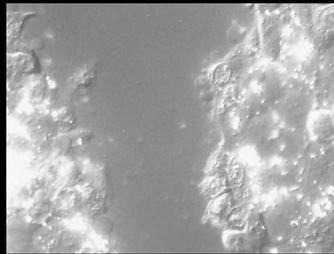
1.5 hr



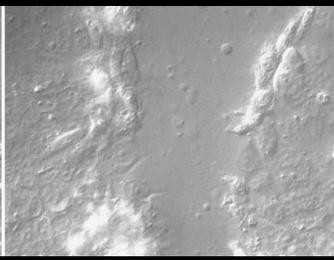
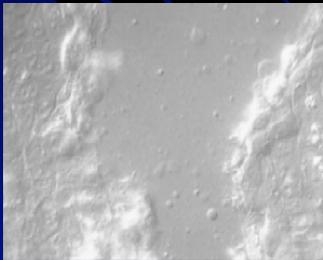
2.0 hr



250 ppm for 2 hr incubation and wash out for microscopy



50 ppm for 12 hr

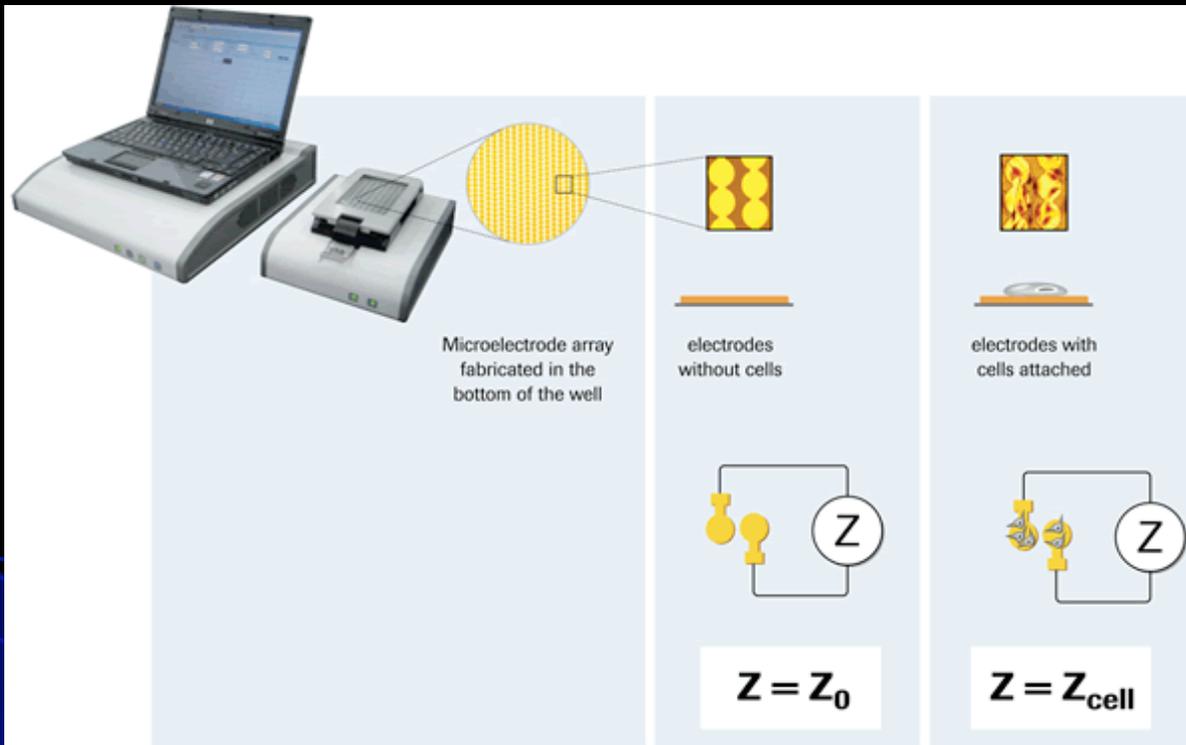


# Scrape Wound Assay to Evaluate Cytotoxicity : Summary

- Scrape wound assay provides a sensitive measure of cellular physiology without the use of fluorescence
- HfO<sub>2</sub> has detrimental effects on cell spreading and/or migration (e.g., cytotoxicity) prior to inducing cell death
- Cellular effects of HfO<sub>2</sub> as low as 50 ppm (lower concentrations not yet tested)
- Results suggest that mechanisms involved in migration (e.g., adhesion) could be cytotoxic targets for HfO<sub>2</sub>
- Scrape wound assay can be difficult to set up and, is limited for high throughput analysis

# Measuring Cytotoxicity Using Impedance

- The xCELLigence System (Roche)

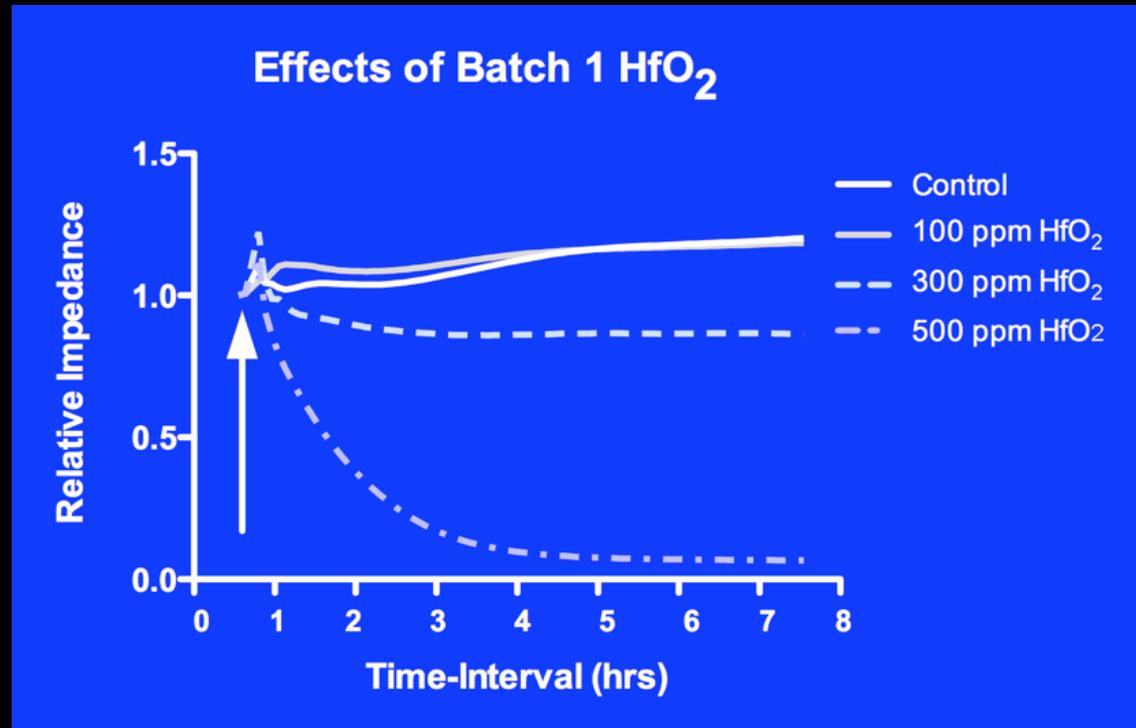


- Uses electrical impedance across interdigitated micro-electrodes integrated on the bottom of tissue culture E-Plates to monitor cellular events in real time without the incorporation of fluorescent labels

- In effect, more cells attached = more impedance
- As cells die (cytotoxicity), lose impedance
- 96 well format allows for relatively high throughput analysis

# Impedance Measurement of Batch-1 360 Cytotoxicity on 16HBE14o- Cells

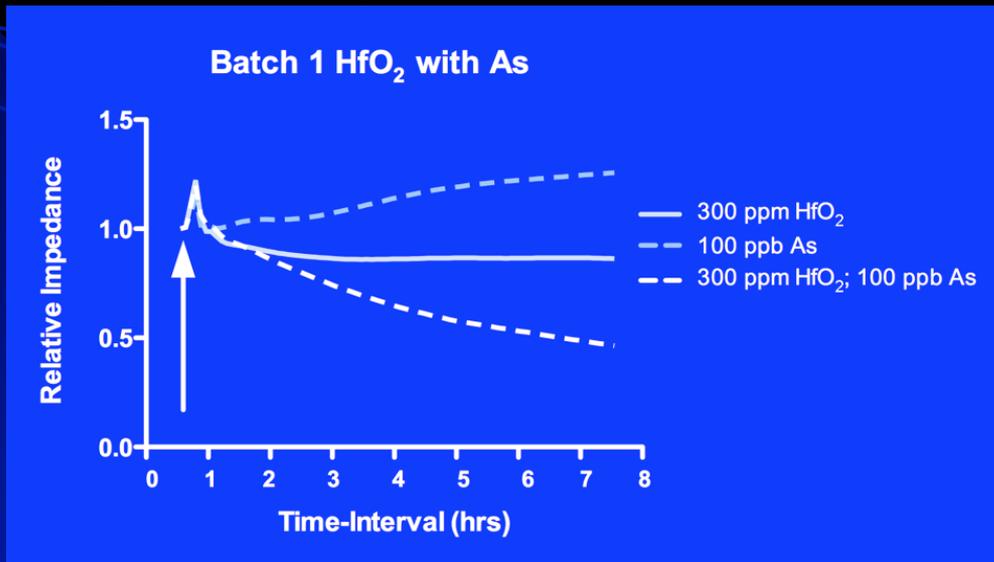
- Grow 16HBE14o-cells on E-Plates
- Add HfO<sub>2</sub> and monitor impedance over time
- Experiments done in series of n=6



- Cells tolerate 100 ppm HfO<sub>2</sub> over the 8 hr period
- 300 ppm and 500 ppm HfO<sub>2</sub> display increasing cytotoxicity
- Note agreement with Live-Dead assay at 2 hrs, but increased cytotoxicity of 500 ppm over time

# Synergistic toxicity of HfO<sub>2</sub> and Arsenic

- Arsenic is a metalloid toxicant that is actively removed from regulated drinking water
- Iron nanoparticles are used to eliminate Arsenic from drinking water
- Arsenic can also be part of materials used in NP production
- Research Question: Does association with HfO<sub>2</sub> lead to cytotoxicity?



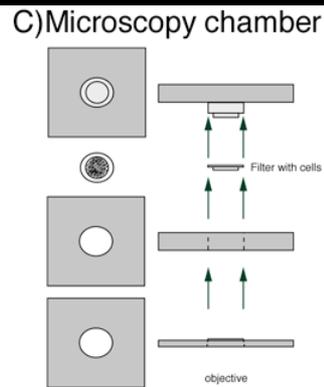
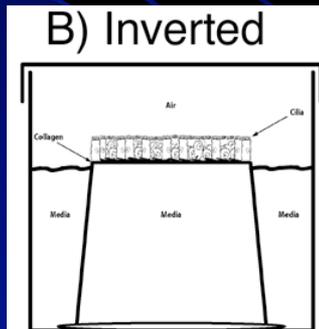
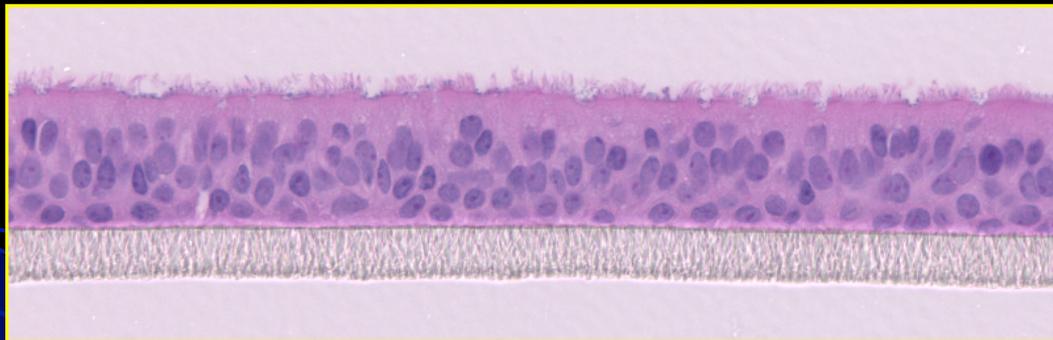
- 100 ppb As is not cytotoxic
- 300 ppm HfO<sub>2</sub> is slightly cytotoxic
- Pre-incubation of 300 ppm HfO<sub>2</sub> with 100 ppb Arsenic significantly increases cytotoxicity

# Summary

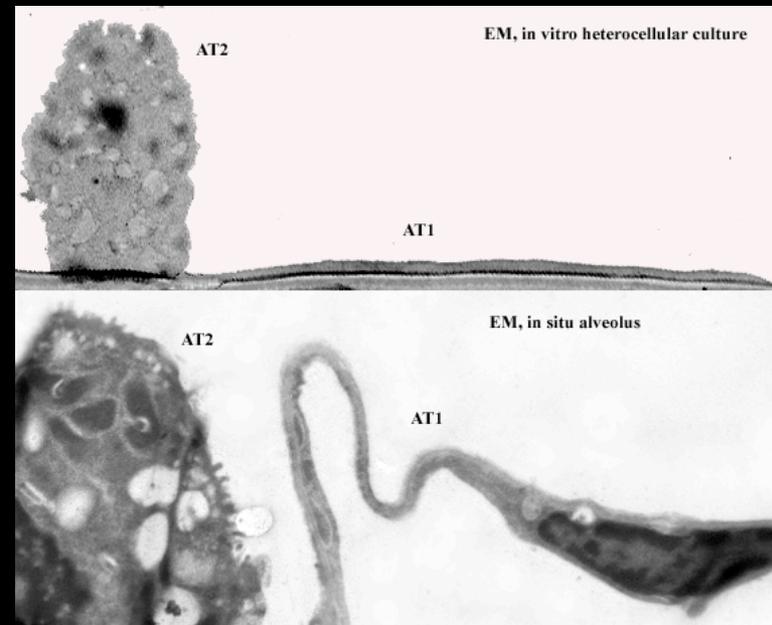
- A mixture of assays can be used to evaluate cytotoxicity
- Size of HfO<sub>2</sub> alone cannot account for HfO<sub>2</sub> cytotoxicity (in agreement with Sierra Group Data)
- Contaminants and or HfO<sub>2</sub> shape/agglomeration can influence cytotoxicity -- how much each of these components contribute to cytotoxicity is unknown
- Considerations for future studies:
  - Immortalized cell models do not necessarily reflect *in vivo* tissue
  - Delivery of nanoparticles to cells not necessarily representative of *in vivo* environment

# Future Directions

- 1) Improved characterization of model nanoparticles
  - 2) Understanding mechanisms of cytotoxicity
  - 3) Improved cell models and delivery
- Normal Human Bronchial Epithelial Cells



## Rat Alveolar Epithelial Cells



# Credits

## Principal Investigators:

Scott Boitano<sup>1</sup>, Reyes Sierra<sup>2</sup>, Jim Field<sup>2</sup>, Buddy Ratner<sup>3</sup>, Farhang Shadman<sup>2</sup>

<sup>1</sup> **Arizona Respiratory Center & Dept. of Physiology  
and Dept Chemical & Environmental Engineering<sup>2</sup>, University of Arizona**

<sup>3</sup> **University of Washington Engineered Biomaterials (UWEB)**

## Student Investigators:

Cara L. Sherwood: Department of Cell Biology and Anatomy, University of Arizona

Nathan Runyan: Department of Physiology, University of Arizona